

# A novel long chain toxin variant from *Androctonus Australis* scorpion venom activates currents carried by $K_v7.4$ channels

E. Miceli<sup>1</sup>, Z. Landoulsi<sup>2</sup>, A. Palmese<sup>3</sup>, A. Amoresano<sup>3</sup>, G. Marino<sup>3</sup>, M. El Ayeb<sup>2</sup>, R. Benkhalifa<sup>2</sup>, M. Tagliatalata<sup>1,4</sup>

<sup>1</sup>Div. Pharmacology, Dept. of Neuroscience, University of Naples Federico II, Naples, Italy

<sup>2</sup>Laboratoire des Venins et Molécules Thérapeutiques, Institut Pasteur de Tunis, Université Tunis-El Manar, Tunis-Belvédère, Tunisia

<sup>3</sup>Dept. of Chemical Sciences, University of Naples Federico II, Naples, Italy

<sup>4</sup>Dept. of Medicine and Health Science, University of Molise, Campobasso, Italy

The  $K_v7$  subfamily of voltage-gated potassium channels ( $K_v7.1$ - $K_v7.5$ ), plays a key pathophysiological role in controlling excitability of cardiac, neuronal, and sensory cells. Mutations in four of five  $K_v7$  genes are responsible for human diseases like cardiac arrhythmia, epilepsy or deafness (Soldovieri et al., 2011). In particular,  $K_v7.4$  channels are expressed in central auditory pathways and in sensory hair cells of the inner ear; mutations in  $K_v7.4$  underlie a rare form of slowly progressive deafness.  $K_v7.4$  channels also regulate vascular (Yeung et al., 2007) and non-vascular smooth muscle contraction, and skeletal muscle differentiation (Iannotti et al., 2013). Drugs acting as openers of  $K_v7$  channels might be effective against neuropsychiatric disorders characterized by neuronal hyperexcitability (including migraine, epilepsy, and neurophatic pain), as well as genitourinary, vascular, and skeletal muscle diseases.

Since scorpion's venoms contain peptides active on sodium and potassium channels, the aim of the present work has been to investigate the *Androctonus australis* (Aa) scorpion venom as a potential source for novel molecular entities active on  $K_v7$  channels.

To this aim, the whole venom from the Aa scorpion was separated by gel filtration chromatography to obtain five partially-resolved fractions; among these, four correspond to non-toxic fractions (M1, M2, M3, and M4), and one (G50) was toxic *in-vivo*. The M2 nontoxic fractions dose-dependently increased  $K_v7.4$  currents in *Xenopus* oocytes and caused a 7-8 mV negative voltage shift in current activation threshold. To identify the active molecule(s) in the M2 fraction, we performed different steps of HPLC purification, obtaining 9 fractions; among these, only the P8 fraction showed  $K_v7.4$ -activating properties. Within this fraction, by means of mass spectral techniques, we identified a novel variant of AaTXK $\beta$ (AaTXK $\beta_{(2-64)}$ ), a toxin belonging to long chain peptides active on  $K^+$  channels ( $\beta$ -KTX), whose sequence had been previously predicted by screening of a venom glands cDNA library from Aa (Legros et al., 1998). This variant differs from the canonical AaTXK $\beta_{(1-64)}$  for the lack of the N-terminal lysine.

To investigate whether AaTXK $\beta_{(2-64)}$  was selective for  $K_v7.4$  subunits, we performed electrophysiological experiment using patch-clamp technique in whole-cell configuration in CHO mammalian cells expressing homomeric  $K_v7.1$ ,  $K_v7.2$ ,  $K_v7.3$ , and  $K_v1.1$  channels, as well as heteromeric  $K_v7.2/3$  channels. Similarly to  $K_v7.4$  channels, aTXK $\beta_{(2-64)}$  significantly increased  $K_v7.3$ ,  $K_v7.2/3$ , and  $K_v7.4$  currents by about 70% (at -40 mV) and caused a 5-10 mV hyperpolarization shift in the activation threshold. By contrast, AaTXK $\beta_{(2-64)}$  failed to modify current amplitudes at depolarized potentials or activation threshold in  $K_v7.1$ ,  $K_v7.2$  and  $K_v1.1$  channels.

In conclusion, in the present study, we have identified and characterized a novel variant of AaTXK $\beta$  from the Aa scorpion acting as a the first natural activator of  $K_v7$  channels. This study may set the basis for developing novel pharmacological tool acting on  $K_v7$  channels.

## Bibliography

Iannotti et al. (2013). *Mol Biol Cell* 24: 274-284

Legros et al. (1998). *FEBS Lett* 431: 375-380

Yeung et al. (2007). *Br J Pharmacol* 151: 758-770

Soldovieri et al. (2011). *Physiology* 26: 365-376